# QUINONES FROM ARDISIA CORNUDENTATA

Z. TIAN, M. N. CHANG\*, M. SANDRINO, L. HUANG, J. X. PAN, B. ARISON, J. SMITH and Y. K. T. LAM Merck Sharp & Dohme Research Laboratories, Dept. of Membrane and Arthritis Research, Rahway, New Jersey 07065, U.S.A.

(Received 21 January 1987)

Key Word Index—Ardista cornudentata; Myrsinaceae; benzoquinone; ardisianone; cornudentanone; leukotriene binding assay; inhibition.

Abstract—The structure of a new benzoquinone, cornudentanone, isolated from Ardisia cornudentata M. was determined.

# INTRODUCTION

Ardisia cornudentata Mez. is one of the many Ardisia species used in folk medicine in the south east part of China. It is used as an anti-inflammatory/analgesic medication and to improve general blood circulation. It is also used as an antidote for snake and insect bites. Like many other Myrsinaceae species, Ardisia is known to contain many benzoquinone pigments [1]. In a study to identify the active components, different extracts of the root of A. cornudentata were evaluated in several in vitro receptor and enzymatic assays in our laboratory. The methylene chloride extract was found to inhibit the binding of leukotriene, D<sub>4</sub>(LTD<sub>4</sub>); to a receptor preparation from guinea pig lung tissue. Using the LTD<sub>4</sub> binding assay as a guide, two benzoquinones (1a and 1b) were obtained as active components and were identified. Ardisianone (1a) is a known compound [2]. The structure assignment of the new benzoquinone cornudentanone (1b) and the binding data for both 1a and 1b are reported.

\*Assignments may be interchanged.

#### RESULTS AND DISCUSSION

Ardisianone (1a) and cornudentanone (1b) accounted for all the  $LTD_4$  receptor antagonistic activity of the crude methylene chloride extract.

Cornudentanone (1b),  $C_{22}H_{34}O_5$ , is a yellow amorphous solid. It accounts for 0.05% by weight of dry roots. Cornudentanone (1b) is optically active,  $[\alpha]_D^{22} = -31.5^\circ$ (CHCl<sub>3</sub>; c 1.60). Its UV spectra (methanol solution) displayed  $\lambda_{\text{max}}$  (log  $\varepsilon$ ); 363 (2.97), 264 (4.02). Its IR (CH<sub>2</sub>Cl<sub>2</sub> solution) absorption bands at 2920, 2850, 1734, 1680, 1627, 1600, 1452, 1370, 1320, 1230 and 1050 cm<sup>-1</sup> are characteristic of a 2,6-di-substituted benzoquinone [6]. Its EI mass spectrum showed fragments at (m/z): 380  $[M+2H]^+$ , 378  $[M]^+$ , 336, 318, 194, 153 and 152. Elemental analysis (calc. for C<sub>22</sub>H<sub>34</sub>O<sub>5</sub>: C, 69.81 % H, 9.05%. Found: C, 69.43% H, 9.37%, confirmed the empirical formula assigned. The <sup>1</sup>H NMR spectrum showed resonances at (CDCl<sub>3</sub>);  $\delta$ 0.91 (t, 3H), 1.28 (m, 18H), 1.58 (m, 2H), 1.95 (s, 3H), 2.43 (ddd, HCH, J = 14.0, 8.8, 1.0 Hz), 2.83 (ddd, HCH, J = 14.0, 3.5, 1.3 Hz), 3.80 (s, 3H) 5.00 (m, 1H), 5.86 (d 1H, J = 2.4 Hz) and 6.43 (ddd, 1H, J = 2.4, 1.3, 1.0 Hz). <sup>13</sup>C NMR (CDCl<sub>3</sub>);  $\delta$ 14.1 (Me), 21.1 (MeCO), 22.7 (C-12'), 25.4 (C-4'), 29.3, 29.4, 29.5, 29.57, 29.6 (C<sub>2</sub>) (C-5'-C-10'), 31.9 (C-11'), 34.2 (C-3')\*, 34.5 (C-1')\*, 56.4 (MeO) 72.3 (HCO), 107.3 (C-3), 134.5 (C-5), 143.5 (C-6), 158.9 (C-2), 171.0 (COO), 181.6 (C-1) and 187.4 (C-4).

Hydrogenation of 1b over platinum oxide in acetic anhydride yielded a colourless oily product, 2. Examination of its <sup>1</sup>H NMR spectrum revealed one additional acetyl group and the compound was identified as 4-O-acetyldihydrocornudentanone (2). The spectral data agreed for the most part with the data reported in ref. [5] for 4-O-acetyldihydroardisianone.

Compound 1a was identified as ardisianone. It accounted for 0.084% by weight of dry roots. Its UV, IR, EIMS and <sup>1</sup>H NMR spectral data were identical to those reported in the literature [2]. Elemental analysis (Calc. for  $C_{24}H_{38}O_5$ : C, 70 90% H, 9.42%. Found. C, 70.28%, H, 9.50%) provided further support for the assignment. Compound 1a is optically active,  $[\alpha]_D^{22} = -40^\circ$  (CHCl<sub>3</sub>; c 1.75).

Cornudentanone (1b) and ardisianone were evaluated in the <sup>3</sup>H-LTD<sub>4</sub>, <sup>3</sup>H-LTC<sub>4</sub> and <sup>3</sup>H-LTB<sub>4</sub> receptor binding assays. Cornudentanone inhibited <sup>3</sup>H-LTB<sub>4</sub> receptor binding in a dose-dependent manner with an 1C<sub>50</sub> of 1.8

<sup>\*</sup>Author to whom correspondence should be addressed.

2362 Z. TIAN et al.

 $\times$  10<sup>-5</sup> M. Cornudentanone is also active in inhibiting the  $^3$ H-LTD<sub>4</sub> binding with an IC<sub>50</sub> of 1.58  $\times$  10<sup>-5</sup> M. Ardisianone also showed inhibition in the  $^3$ H-LTB<sub>4</sub> binding assay in a dose-dependent manner with an IC<sub>50</sub> of 2.7  $\times$  10<sup>-5</sup> M.

## **EXPERIMENTAL**

<sup>1</sup>H and <sup>13</sup>C NMR: 400 MHz, TMS as an int. standard.

Isolation of cornudentanone (1b) and ardisianone (1a). The dried plant material was collected from Taiwan. Twenty grams of the roots of A. cornudentata was ground and macerated in 500 ml CH<sub>2</sub>Cl<sub>2</sub> for 24 hr at ambient temp. The extract was filtered and concentrated under red. pres. to yield 0.25 g of a brown semisolid. This crude extract was found active in the LTD, receptor binding assay with  $1C_{50}$  of  $3.5 \times 10^{-5}$  M. A flash column (2  $\times$  30 cm) packed with 30 g of silica gel (Kieselgel 60, 230-400 mesh) was equilibrated with hexane-EtOAc (20:1). The crude mixture from the CH2Cl2 extract was dissolved in 0.5 ml CH<sub>2</sub>Cl<sub>2</sub> was loaded onto the column and eluted at 6 psi of N<sub>2</sub> with 150 ml hexane-EtOAc (20:1), 150 ml hexane-EtOAc (10:1), 150 ml hexane-EtOAc (5:1), 150 ml hexane-EtOAc (1:1) and finally 150 ml EtOAc-MeOH (1:1). Fractions of 30 ml were collected and concentrated. Samples from each of the 25 fractions collected were tested in the LTD4 receptor binding assay. Active fractions were combined to give 34 mg of a bright yellow semisolid. This fraction was further purified by semi-prep. HPLC using a normal phase silica column (Zorbax SIL, 0.46 × 25 cm) and eluted with hexane-EtOAc (19:3). Two compounds active in LTD<sub>4</sub> binding assay were obtained: cornudentanone (1b) and ardisianone (1a).

Reduction of cornudentanone (1b). Cornudentanone (5 mg, 0.013 mmol.) was dissolved in 0.5 ml HOAc and 0.1 ml  $Ac_2O$ . Five milligrams of  $PtO_2$  was added and the mixture was hydrogenated under 40 psi of  $H_2$  on a PAR shaker for 15 min. Ten ml of EtOAc was added and the catalyst filtered off. The resulting soln was concentrated and residual solvent removed under red. pres. A colourless oily product (5.2 mg, 90% yield) was

recovered and was identified as 4-acetyldihydrocornudentanone

LTB<sub>4</sub>, LTC<sub>4</sub> and LTD<sub>4</sub> binding assays. Guinea pig lung membranes were prepared by the procedure of ref. [4]. LTC<sub>4</sub> and LTD<sub>4</sub> binding assays were performed by the procedure of ref. [5] with a minor modification. Routinely, 0.80 nM [<sup>3</sup>H] LTD<sub>4</sub> or 0.40 nM [<sup>3</sup>H] LTC<sub>4</sub> was used as the ligand. After incubation at room temp. for 1 h, bound radioactivity was harvested with GF/B filters using the Skatron cell harvester. The amount of radioactivity remaining on the filters was determined in Scintin Verse 1 Cocktail.

[3H] LTB<sub>4</sub> receptor binding assay was run with human PMN membranes. Human PMN leukocytes were isolated from human leukocyte concentrate [6, 7]. Membranes were prepared by the sonification and centrifugation of human PMN leukocytes. Routinely, 70 pM of [3H]-LTB<sub>4</sub> was used as the ligand. Specific binding and nonspecific binding was defined as the difference between total binding and binding in the presence of 0.65  $\mu$ M LTB<sub>4</sub> [8]. All assays were run in triplicate and experiments were repeated twice.

## REFERENCES

- 1. Ogawa, H. and Natori, S. (1968) Phytochemistry 7, 773.
- Kusumi, T., Kakisawa, H., Chen, Y. P., Lyn, Y.- I. and Hsu, H.-Y. (1978) Bull. Chem. Soc. Japan 51, 943.
- Natori, S., Nishikawa, H. and Ogawa, H. (1964) Chem. Pharm. Bull. 12, 236.
- Pong, S. S. and Dehaven, R. N. (1983) Proc. Natl. Acad. Sci. U.S.A. 80, 7415.
- Pong, S. S., Dehaven, R. N., Kuehl, F. A. Jr and Egan, R. W. (1983) J. Biol. Chem. 258, 9616.
- English, D. and Anderson, B. R. (1974) J. Immunol. Methods 5, 249.
- Rollins, T. E. and Springer, M. S. (1985) J. Biol. Chem. 260, 7157
- Goldman, D. W. and Goetzl, E. J. (1984) J. Exp. Med. 159, 1027.